Contents

| List of Tables 9 | | | | | | | |
|------------------|------------|---|--|--|--|--|--|
| Li | st of | Pigures | 11 | | | | |
| 1 | Intr | ntroduction | | | | | |
| 2 | 2.1 2.2 | Particle of literatures Faxonomy of Equid Herpesviruses | 16 16 16 | | | | |
| | | All Morphological structure 2.2.2 Genome structure 2.2.3 The DNA polymerase (open reading frame 30) 2.2.4 Latency 2.2.5 Epidemiology 2.2.6 Clinical signs 2.2.7 Treatment 2.2.8 Prophylaxis and Management 2.2.9 Pathogenesis | 16 17 17 18 19 20 21 22 22 | | | | |
| 3 | Obj | 2.2.10 Detection methods | 27 29 | | | | |
| 4 | Mat | rial and methods | 30 | | | | |
| | 4.1 | Proof of material . 1.1.1 Chemicals and solutions . 1.1.2 Materials . 1.1.3 Equipment . EHV-1 and -4 reference strains . 1.2.1 EHV-1 reference strains . 1.2.2 EHV-4 reference strains . | 30 30 31 31 33 33 33 | | | | |
| | 4.3 | Wild animal and cattle EHV strains | 34 34 35 | | | | |
| | 4.4 | Sample origin 4.4.1 Single abortion cases 4.4.2 Abortion outbreaks in Germany 4.4.3 Abortion outbreak stud farm 1 4.4.4 Abortion outbreak stud farm 2 4.4.5 Abortion outbreak stud farm 3 4.4.6 Abortion outbreak stud farm 4 1.4.7 Neurological cases | 35 36 36 36 37 37 38 39 | | | | |
| | 4.5 | Cell cultures | 40 40 | | | | |

| 4.6 | Serological tests | 41 |
|------|--|----|
| | ` ' | 41 |
| | 4.6.2 Immunofluorescence assay (IFA) | 42 |
| 4.7 | PBMC isolation from citrated blood | 43 |
| 4.8 | DNA preparation | 44 |
| | 4.8.1 DNA preparation from tissue samples and PBMCs | 44 |
| | 4.8.2 Viral DNA preparation from virus stocks, infected cell culture supernatants, | |
| | nasal swabs, lung fluids, amniotic fluid and semen | 44 |
| 4.9 | Analytical gel electrophoresis | 45 |
| 4.10 | | 46 |
| | 4.10.1 Gel electrophoresis | 46 |
| | 4.10.2 Photometry | 46 |
| 4.11 | • | 46 |
| | 4.11.1 ORF 30 nested PCR | 47 |
| | | 49 |
| | | 50 |
| 4.12 | | 50 |
| | , | 50 |
| | • | 51 |
| 4.13 | • | 51 |
| 4.14 | Statistics | 52 |
| | | |
| | | 53 |
| 5.1 | | 53 |
| | 5.1.1 Establishment of the ORF 30 nested PCR with selected EHV-1/-4 reference | |
| | | 54 |
| | • | 55 |
| 5.2 | Test of the restriction enzyme analysis Sal I with the ORF 30 fragments of the | |
| | | 56 |
| 5.3 | Review of the ORF 30 fragments of the selected EHV-1 reference strains by sequencing | 58 |
| | 5.3.1 Chromatograms of the ORF 30 fragments of the selected EHV-1 reference | |
| | | 59 |
| 5.4 | Occurrence of the non-neuropathogenic versus the neuropathogenic genotype in | |
| | | 59 |
| | | 61 |
| | | 62 |
| 5.5 | | 64 |
| 5.6 | Sample character of the EHV-1 abortion cases | 65 |
| 5.7 | Occurrence of the non-neuropathogenic versus the neuropathogenic genotype in | |
| | EHV-1 abortion cases | 65 |
| | · · | 65 |
| | | 67 |
| | 5.7.3 Sequencing of selected ORF 30 amplicons | 74 |

| | | 5.7.4 | Additional initiation in the OKF 30 amplicons from abortion cases | 18 |
|----|------|----------------|--|--------------------|
| | | 5.7.5 | Stud farm 4: Detection of viral DNA in PBMC and determination of the | |
| | | | genotype | 78 |
| | 5.8 | Serolo | gical studies | 79 |
| | | 5.8.1 | Stud farm 3 with abortions and neurological signs | 79 |
| | | 5.8.2 | Stud farm 4 with an abortion outbreak | 83 |
| 6 | Disc | cussion | 1 | 85 |
| | 6.1 | Detect | sion of the non-neuropathogenic versus the neuropathogenic EHV-1 genotype | |
| | | in abo | rtion cases in Germany | 85 |
| | 6.2 | EHV-1 | l in stallions | 89 |
| | 6.3 | A dditi | onal nucleotide exchanges in the ORF 30 in Germany | 90 |
| | | 6.3.1 | Nucleotide exchange at position 2258 | 90 |
| | | 6.3.2 | Nucleotide exchange at position 2269 | 91 |
| | 6.4 | | tion of the neuropathogenic genotype in cattle and archived wild equid strains | 91 |
| | | 6.4.1 | Nucleotide exchange at position 2262 | 92 |
| | 6.5 | | cion of the neuropathogenic EHV-1 genotype in neurological cases in Germany | 92 |
| | 6.6 | | gical and molecular biological detection of EHV-1 in abortion outbreaks on | |
| | | | ud farms | 93 |
| | | 6.6.1 | Stud farm 3 | 93 |
| | | 6.6.2 | Stud farm 4 | 94 |
| 7 | Sun | nmary | | 96 |
| 8 | Zusa | ammer | nfassung | 97 |
| 9 | Bibl | liograp | bhy | 98 |
| 10 | Ann | | | 10 |
| 10 | | | | 1 10 110 |
| | 10.1 | | once strains Origin of the EHV-1/-4 reference strains and wild animal strains, sample | 110 |
| | | 10.1.1 | in the second of | 110 |
| | | 10 1 2 | Results of the ORF 30 nested PCR, SalI restriction enzyme analysis and | 110 |
| | | 10.1.2 | | 111 |
| | | 10.1.3 | | 111 |
| | 10.2 | | • | 112 |
| | 20.2 | | Origin of the 67 EHV-1 abortion cases, sample character, processed sam- | |
| | | | ple character, ORF 30 nested PCR and Sal I restriction enzyme results. | |
| | | | Selection of the ORF 30 amplicons for sequencing | 112 |
| | | 10.2.2 | Sequences of the abortion strains from 1987 to 2009 | |
| | 10.3 | | - | 121 |
| | | | Results of the ORF 30 nested PCR, SalI restriction enzyme analysis and | |
| | | | selection of the ORF 30 amplicons for sequencing of neurological cases | 121 |
| | | 10.3.2 | Sequences of the neurological EHV strains | |
| | 10.4 | Repor | t of the clinical signs of the sampled horses from stud farm 3 | 121 |

| 10.5 | Serology | 122 |
|------|--|-----|
| | 10.5.1 Stud farm 3: EHV-1/-4 serum neutralization and immunofluorescence assay | 122 |
| | 10.5.2 Stud farm 4: EHV-1/-4 neutralization test | 123 |
| 10.6 | List of own publications | 124 |
| 10.7 | Danksagung | 125 |
| 10.8 | Selbständigkeitserklärung | 126 |